

FEMALE ARCHESPORIUM OF ANGIOSPERMS: STRUCTURE, EVOLUTION, AND BIOLOGICAL SIGNIFICANCE

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The review is devoted to the formation of unicellular and multicellular archesporium in female reproductive structures (ovules), differentiation of its cells into megasporocytes, and regulation of these processes. The terminology used in relation to the archesporium is considered, various points of view on the archesporium nature, its place and time of initiation in the ovule and transition into megasporocytes have been analyzed. Based on this, it was concluded that at the early stages of the ovule development, all its cells are identical and meristematic; later, one or more cells in the subepidermal or deeper layers start to differentiate as archesporial (sporogenous) cells, showing differences from other nucellar cells in terms of ultrastructural organization, composition cell wall, characteristics of chromatin in the nucleus.

The differentiation of unicellular and, apparently, multiple archesporium is probably timed to the initiation of the inner integument and is regulated by the expression of the *WUSHEL* (*WUS*), *NOZZLE* (*NZZ*), *WINDHOSE1* (*WIH1*), and *WIH2* genes responsible for cell differentiation in the ovule primordium, and *AINTEGUMENTA* (*ANT*), *BEL1*, *INNER NO OUTER* (*INO*), responsible for the integument initiation. Possible factors leading to the differentiation of several sporogenous cells in the ovule of plants, which are typically characterized by unicellular archesporium (*Arabidopsis*, maize, rice), are considered. These include: mutations in the genes that regulate the differentiation of sporogenous cells; disturbances in epigenetic regulation including the action of small RNAs, DNA methylation and cell cycle mechanisms; as well as irregularities in spatio-temporal coordination of the processes of the ovule development and positional information.

The role of callose in the determination of only one (rarely two) megasporocyte in a multicellular sporogenous complex for its entry into meiosis and the formation of a single gametophyte in the ovule is indicated. It has been suggested that callose not only isolates the megasporocyte for megasporogenesis, but creates a block for the transfer of signaling molecules to neighboring megasporocytes located on the periphery of the sporogenous complex, which stops their development.

The data of the analysis of the evolution of the female archesporium are presented. It has been suggested that the multiple archesporium derived from the unicellular one in the historical development of angiosperms. This is evidenced by the analysis of the distribution of different types of the archesporium among the orders on the APG IV phylogenetic system (2016). The orders of Basal Dicots and Monocots, located at the base of the phylogenetic tree, are characterized by unicellular archesporia (excluding Laurales). The multiple archesporium is characteristic, as a rule, of more advanced orders of Eudicots, but even in these orders and their families, the multiple archesporium does not dominate and appears in fewer species than unicellular one. Sporadic appearance of additional archesporial cells (usually not developing further) in species with the unicellular archesporium, as well as their appearance as a result of mutations, may indicate the secondary nature of multiple archesporium. Probably, multiple archesporium originated as a result of adaptation to preserve reproductive potential of plants, when barriers to the development of a sporogenous cell appear or its frequent death that, apparently, is its biological role.

Keywords: ovule, archesporium, multiple archesporium, sporogenous cell, megasporocyte, megasporogenesis, evolution of the archesporium

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REFERENCES

- APG IV. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. — *Bot. J. Linn. Soc.* 181 (1): 1–20.
- Armenta-Medina A., Demesa-Arévalo E., Vielle-Calzada J.P. 2011. Epigenetic control of cell specification during female gametogenesis. — *Sex. Plant Reprod.* 24 (2): 137–147.
- Bai S.N. 2017. Two types of germ cells, the sexual reproduction cycle, and the double-ring mode of plant developmental program. — *Plant Signaling & Behavior*. 12 (7): e1320632.
- Bajon C., Horlow C., Motamayor J., Sauvanet A., Robert D. 1999. Megasporogenesis in *Arabidopsis thaliana* L.: an ultrastructural study. — *Sex. Plant Reprod.* 12 (2): 99–109.
- Balasubramanian S., Schneitz K. 2000. NOZZLE regulates proximal-distal pattern formation, cell proliferation and early sporogenesis in *Arabidopsis thaliana*. — *Development*. 127 (19): 4227–4238.
- Batygina T.B. 2002. Ovule and seed viewed from reliability of biological systems. — In: *Embryology of flowering plants. Terminology and concepts. Vol. 1. Generative organs of flower.* Enfield (NH, USA). P. 214–217.
- Batygina T.B. 2014. *Biology of plant development.* Life Symphony. Saint Petersburg. 764 p. (In Russ.).
- Bencivenga S., Colombo L., Masiero S. 2011. Cross talk between the sporophyte and the megagametophyte during ovule development. — *Sex. Plant Reprod.* 24 (2): 113–121.
- Bencivenga S., Simonini S., Benkova E., Colombo L. 2012. The transcription factors BEL1 and SPL are required for cytokinin and auxin signaling during ovule development in *Arabidopsis*. — *Plant Cell*. 24 (7): 2886–2897.
- Berger F., Twell D. 2011. Germline specification and function in plants. — *Annual Review of Plant Biology*. 62: 461–484.
- Berger X. 1953. Untersuchungen über Embryologie partiell apomiktischer Rubusbastarde. — *Berichte der Schweizerischen Botanischen Gesellschaft*. 63: 224–266.
- Bichet A., Desnos T., Turner S., Grandjean O., Höfte H. 2001. BOTERO1 is required for normal orientation of cortical microtubules and anisotropic cell expansion in *Arabidopsis*. — *Plant J.* 25 (2): 137–148.
- Bloemendal S., Kuck U. 2013. Cell-to-cell communication in plants, animals, and fungi: a comparative review. — *Naturwissenschaften*. 100 (1): 3–19.
- Bouman F. 1984. *The ovule.* — *Embryology of angiosperms.* Berlin. P. 123–157.
- Brukhin V. 2017. Molecular and genetic regulation of apomixis. — *Russian Journal of Genetics*. 53 (9): 1001–1024.
- Burk D.H., Ye Z.H. 2002. Alteration of oriented deposition of cellulose microfibrils by mutation of a katanin-like microtubule-severing protein. — *Plant Cell*. 14 (9): 2145–2160.
- Burton R.A., Gidley M.J., Fincher G.B. 2010. Heterogeneity in the chemistry, structure and function of plant cell walls. — *Nature Chemical Biology*. 6 (10): 724–732.
- Cao L., Wang S., Venglat P., Zhao L., Cheng Y., Ye S., Qin Y., Datla R., Zhou Y., Wang H. 2018. Arabidopsis ICK/KRP cyclin-dependent kinase inhibitors function to ensure the formation of one megaspore mother cell and one functional megaspore per ovule. — *Plos Genetics*. 14 (3): e1007230.
- Carpita N., Sabulase D., Montezinos D., Delmer D.P. 1979. Determination of the pore size of cell walls of living plant cells. — *Science*. 205 (4411): 1144–1147.
- Ceccato L., Masiero S., Roy D.S., Bencivenga S., Roig-Villanova I., Ditengou F.A., Palme K., Simon R., Colombo L. 2013. Maternal control of PIN1 is required for female gametophyte development in *Arabidopsis*. — *PLoS One*. 8 (6): e66148.
- Cecchi Fiordi A., Mariotti Lippi M., Marini S., Tani G. 1996. Ultrastructural features of megasporogenesis in *Torreya nucifera* (Taxaceae). — *Plant Systematics and Evolution*. 202 (1/2): 13–25.
- Cheng C.Y., Mathews D.E., Schaller G.E., Kieber J.J. 2013. Cytokinin-dependent specification of the functional megaspore in the *Arabidopsis* female gametophyte. — *Plant J.* 73 (6): 929–940.
- Chevalier É., Loubert-Hudon A., Zimmerman E.L., Matton D.P. 2011. Cell-cell communication and signaling pathways within the ovule: from its inception to fertilization. — *New Phytologist*. 192 (1): 13–28.
- Clark S.E. 2001. Cell signalling at the shoot meristem. — *Nature Reviews Molecular Cell Biology*. 2 (4): 276–284.
- Comparative embryology of flowering plants. 1981, 1983, 1985, 1987, 1990. Vol. 1–5. Leningrad (In Russ.).
- Crepet W.L., Delevoryas T. 1972. Investigations of North American cycadeoids: Early ovule ontogeny. — *Am. J. Bot.* 59 (2): 209–215.
- Czapik R. 1961–1962. Embryological studies in the genus *Potentilla* L. I. *P. crantzii*. — *Acta Biologica Cracoviensia, series Botanica*. 14: 97–119.
- Dahlgren K.V.O. 1927. Die Morphologie des Nuzellus mit besonderer Berücksichtigung der deckzellosten Typen. — *Jahrbücher für Wissenschaftliche Botanik*. 67: 347–426.
- Davis G.L. 1966. *Systematic embryology of the angiosperms.* New York, London, Sydney. 528 p.
- Dawe R.K., Freeling M. 1992. The role of initial cells in maize anther morphogenesis. — *Development*. 116 (4): 1077–1085.
- Dickinson H.G., Grant-Downton R. 2009. Bringing the generation gap: flowering plant gametophytes and animal germlines reveal unexpected similarities. — *Biol. Rev.* 84 (4): 589–615.
- Dupler A.W. 1917. The gametophyte of *Taxus canadensis* Marsh. — *Botanical Gazette*. 64 (2): 115–136.

- Endress P.K. 2011. Angiosperm ovules: diversity, development, evolution. — *Ann. Bot.* 107 (9): 1465–1489.
- Fagerlind F. 1937. Embryologische, zytologische und bestäubungsexperimentelle Studien in der Familie Rubiaceae nebst Bemerkungen über einige Polyploiditätsprobleme. — *Acta Horti Bergiani*. 2: 196–470.
- Fan X., Yuan D., Tang J., Tian X., Zhang L., Zou F., Tan X. 2015. Sporogenesis and gametogenesis in Chinese chinquapin (*Castanea henryi* (Skam) Rehder et Wilson) and their systematic implications. — *Trees*. 29 (6): 1713–1723.
- Friedman W.E., Carmichael J.S. 1998. Heterochrony and developmental innovation: evolution of female gametophyte ontogeny in *Gnetum*, a highly apomorphic seed plant. — *Evolution*. 52 (4): 1016–1030.
- Gerassimova-Navashina E.N. 1958. O gametofite i ob osnovnykh chertakh razvitiya i funktsionirivaniya vosproizvodyaschikh elementov u pokrytosemennykh rasteniy [About gametophyte and main signs of development and functioning reproductive elements in Angiosperms]. — *Problemy botaniki*. 3: 125–167 (In Russ.).
- Goebel K. 1880. Beitrage zur vergleichenden Entwicklungsgeschichte der Sporangien. — *Botanische Zeitung*. 38: 545–553.
- Goebel K. 1933. Organographie der Pflanzen, insbesondere der Archegoniaten und Samenpflanzen. Auf. 3. Teil. 2, 3. Jena. 642 s.
- Gross-Hardt R., Laux T. 2003. Stem cell regulation in the shoot meristem. — *Journal of Cell Science*. 116 (9): 1659–1666.
- Gross-Hardt R., Lenhard M., Laux T. 2002. WUSCHEL signaling functions in interregional communication during *Arabidopsis* ovule development. — *Genes and Development*. 16 (9): 1129–1138.
- Grossniklaus U. 2011. Plant germline development: a tale of cross-talk, signaling, and cellular interactions. — *Sex. Plant Reprod.* 24 (2): 91–95.
- Hake S., Char B.R. 1997. Cell-cell interactions during plant development. — *Genes and Development*. 11: 1087–1097.
- Hayashi K., Surani M.A. 2009. Resetting the epigenome beyond pluripotency in the germline. — *Cell Stem Cell*. 4 (6): 493–498.
- Hernandez-Lagana E., Rodriguez-Leal D., Lua J., Vielle-Calzada J.P. 2016. A multigenic network of ARGONAUTE4 clade members controls early megaspore formation in *Arabidopsis*. — *Genetics*. 204 (3): 1045–1056.
- Hernandez-Lagana E., Mosca G., Mendocilla-Sato E., Pires N., Frey A., Giraldo-Fonseca A., Michaud C., Grossniklaus U., Hamant O., Godin C., Boudaoud A., Grimanelli D., Autran D., Baroux C. 2021. Organ geometry channels reproductive cell fate in the *Arabidopsis* ovule primordium. — *eLife*. 10: e66031.
- Hernandez-Lagana E., Autran D. 2020. H3.1 eviction marks female germline precursors in *Arabidopsis*. — *Plants*. 9 (10): 1322.
- Hjelmqvist H. 1962. The embryo sac development of some *Cotoneaster* species. — *Botaniska Notiser*. 115: 209–236.
- Huala E., Sussex I.M. 1993. Determination and cell interactions in reproductive meristems. — *Plant Cell*. 5 (10): 1157–1165.
- Jiang T., Zheng B. 2022. Epigenetic regulation of megaspore mother cell formation. — *Frontiers in Plant Science*. 12: article 826871.
- Kamelina O.P. 2000. Embryological features in phylogenetic systematics of flowering plants. — *Bot. Zhurn.* 85 (7): 22–33 (In Russ.).
- Kamelina O.P. 2009. Systematic embryology of flowering plants. Vol. 1. Dicotyledons. Barnaul. 501 p. (In Russ.).
- Khan R. 1942. A contribution to the embryology of *Jussiaea repens* L. — *Journal of the Indian Botanical Society*. 21: 267–282.
- Kim I., Kobayashi K., Cho E., Zambryski P.C. 2007. Regulation of plant intercellular communication via plasmodesmata. — In: *Genetic Engineering. Principle and Methods*. Vol. 28. New York. P. 1–15.
- Koltunow A.M. 1993. Apomixis: embryo sacs and embryos formed without meiosis or fertilization in ovules. — *Plant Cell*. 5 (10): 1425–1437.
- Koltunow A.M., Grossniklaus U. 2003. Apomixis: a developmental perspective. — *Annual Review of Plant Biology*. 54: 547–574.
- Konar R.N., Oberoi Y.P. 1969. Recent work on reproductive structure of living conifers and taxads: a review. — *Botanical Review*. 35 (2): 89–116.
- Kordyum E.L. 1978. Evolutionary cytoembryology of angiosperms. Kiev. 219 p. (In Russ.).
- Lee J.Y., Lu H. 2011. Plasmodesmata: the battleground against intruders. — *Trends in Plant Science*. 16 (4): 201–210.
- Leszczuk A., Domaciuk M., Szczuka E. 2018. Unique features of the female gametophyte development of strawberry *Fragaria x ananassa* Duch. — *Scientia Horticulturae*. 234: 201–209.
- Lieber D., Lora J., Schrempf S., Lenhard M., Laux T. 2011. *Arabidopsis WIH1* and *WIH2* genes act in the transition from somatic to reproductive cell fate. — *Current Biology*. 21 (12): 1009–1017.
- Lodkina M.M. 1971. O ponyatii “archesporiya” [On the term “archesporium”]. — *Proceedings of V All-Union Session on the Plant Embryology*. Kishinev. P. 101–102 (In Russ.).
- Lora J., Herrero M., Tucker M.R., Hormaza J.I. 2017. The transition from somatic to germline identity shows conserved and specialized features during angiosperm evolution. — *New Phytologist*. 216 (2): 495–509.
- Lora J., Yang X., Tucker M.R. 2019. Establishing a framework for female germline initiation in the plant ovule. — *Journal of Experimental Botany*. 70 (11): 2937–2949.
- Lucas W.J., Ham B.K., Kim J.Y. 2009. Plasmodesmata — bridging the gap between neighboring plant cells. — *Trends in Cell Biology*. 19 (10): 495–503.
- Mahashwari P. 1950. An introduction to the embryology of angiosperms. New York. 455 p.
- Matzke M.A., Mosher R.A. 2014. RNA-directed DNA methylation: an epigenetic pathway of increasing complexity. — *Nature Reviews Genetics*. 15 (6): 394–408.
- Mendes M.A., Petrella R., Cucinotta M., Vignati E., Gatti S., Pinto S.C., Bird D.C., Gregis V., Dickinson H., Tucker M.R., Colombo L. 2020. The RNA-dependent DNA methylation pathway is required to restrict *SPO-ROCYTELESS/NOZZLE* expression to specify a single

- female germ cell precursor in *Arabidopsis*. – Development. 147 (23): dev194274.
- Naumova T.N. 1987. Family Celastraceae. – Comparative embryology of flowering plants. Leningrad. P. 49–54 (In Russ.).
- Newbigin E., Bacic A., Read S. 2009. Callose and its role in pollen and embryo sac development in flowering plants. – In: Chemistry, Biochemistry and Biology of (1–3)- β -Glucans and Related Polysaccharides. New York, London, Oxford, San Diego. P. 465–498.
- Nikiticheva Z.I., Shamrov I.I. 2002. Archesporium. – Embryology of flowering plants. Terminology and concepts. Vol. 1. Generative organs of flower. Enfield (NH, USA). P. 112–114.
- Nonomura K.I. 2018. Small RNA pathways responsible for non-cell-autonomous regulation of plant reproduction. – Plant Reprod. 31 (1): 21–29.
- Nonumura K., Mioshi K., Eiguchi M., Suzuki T., Miyao A., Hirochika H., Kurata N. 2003. The *MSP1* gene is necessary to restrict the number of cells entering into male and female sporogenesis and to initiate anther wall formation rice. – Plant Cell. 15 (8): 1728–1739.
- Olmedo-Monfil V., Duran-Figueroa N., Arteaga-Vazquez M., Demesa-Arevalo E., Autran D., Grimanelli D., Slotkin R.K., Martienssen R.A., Vielle-Calzada J.P. 2010. Control of female gamete formation by a small RNA pathway in *Arabidopsis*. – Nature. 464: 628–632.
- Pagnussat G.C., Alandete-Saez M., Bowman J.L., Sundaresan V. 2009. Auxin-dependent patterning and gamete specification in the *Arabidopsis* female gametophyte. – Science. 324 (5935): 1684–1689.
- Petrasek J., Mravec J., Bouchard R., Blakeslee J.J., Abas M., Seifertova D., Wisniewska J., Tadele Z., Kubes M., Covanova M., Dhonukshe P., Skupa P., Benková E., Perry L., Krecek P., Lee O.R., Fink G.R., Geisler M., Murphy A.S., Luschnig C., Zazimalová E., Friml J. 2006. PIN proteins perform a rate-limiting function in cellular auxin efflux. – Science. 312 (5775): 914–918.
- Pinto S.C., Mendes M.A., Coimbra S., Tucker M.R. 2019. Revisiting the female germline and its expanding toolbox. – Trends in Plant Science. 24 (5): 455–467.
- Poddubnaya-Arnoldi V.A. 1976. Tsitoembriologiya pokrytosemennykh rasteniy. Osnovy i perspektivy [Cytoembryology of angiosperms. Basis and perspectives]. Moscow. 508 p. (In Russ.).
- Poethig S. 1989. Genetic mosaics and cell lineage analysis in plants. – Trends in Genetics. 5 (8): 273–277.
- Rodkiewicz B., Bednara J. 1976. Cell wall ingrowth and callose distribution in megasporogenesis in some Orchidaceae. – Phytomorphology. 26: 276–281.
- Rodkiewicz B., Bednara J. 2002. Megasporogenesis. – In: Embryology of flowering plants. Terminology and concepts. Vol. 1. Generative organs of flower. Enfield (NH, USA). P. 114–115.
- Rodkiewicz B. 1970. Callose in cell walls during megasporogenesis in angiosperms. – Planta. 93 (1): 39–47.
- Romanov I.D. 1954. Embriologicheskie issledovaniya hlopchatnika. I. Razvitie sporoobrazuyuschih kletok v semyazchatke [Embryological studies of cotton. I. Spore-formed cells development in the ovule]. – Proceeding Middle-Asian State University. 53: 3–58 (In Russ.).
- Savidan Y. 2000. Apomixis: genetics and breeding. – In: Plant Breeding Reviews. New York. P. 13–86.
- Schmidt A., Schmid M.W., Grossniklaus U. 2015. Plant germline formation: common concepts and developmental flexibility in sexual and asexual reproduction. – Development. 142 (2): 229–241.
- Schnarf K. 1927–1929. Embryologie der Angiospermen. Berlin. 689 s.
- Schnarf K. 1931. Vergleichende Embryologie der Angiospermen. Berlin. 354 s.
- Sehgal A., Khurana J.P., Sethi M., Ara H. 2011. Occurrence of unique three-celled megagametophyte and single fertilization in an aquatic angiosperm – *Dalzellia zeylanica* (Podostemaceae–Tristichioideae). – Sex. Plant Reprod. 24 (3): 199–210.
- Shamrov I.I. 1997. Ovule and seed development in *Paeonia lactiflora* (Paeoniaceae). – Bot. Zhurn. 82 (6): 24–46 (In Russ.).
- Shamrov I.I. 2008. Ovule of flowering plants: structure, functions, origin. Moscow. 350 p. (In Russ.).
- She W., Grimanelli D., Rutowicz K., Whitehead M.W., Puzio M., Kotlinski M., Jerzmanowski A., Baroux C. 2013. Chromatin reprogramming during the somatic-to-reproductive cell fate transition in plants. – Development. 140 (9): 4008–4019.
- Sheridan W.F., Avalkina N.A., Shamrov I.I., Batygina T.B., Golubovskaya I.N. 1996. The *mac1* gene: Controlling the commitment to the meiotic pathway in maize. – Genetics. 142 (3): 1009–1020.
- Sieber P., Gheyselinck J., Gross-Hardt R., Laux T., Grossniklaus U., Schneitz K. 2004. Pattern formation during early ovule development in *Arabidopsis thaliana*. – Developmental Biology. 273 (2): 321–334.
- Sladkov A.N., Grevtsova N.A. 1988. O formirovanii stenki mikrosporangiya pokrytosemennykh [On the formation of microsporangium wall in angiosperms]. – Byulleten Moskovskogo Obshchestva Ispytateley Prirody. Otdelenie Biologicheskoe. 93 (4): 69–74 (In Russ.).
- Sladkov A.N., Grevtsova N.A. 1989. O formirovanii stenki megasporangiya pokrytosemennykh [On the formation of megasporangium wall in angiosperms]. – Byulleten Moskovskogo Obshchestva Ispytateley Prirody Otdelenie Biologicheskoe. 94 (3): 75–79 (In Russ.).
- Solntseva M.P. 1965. On the development of the multicellular archesporium in strawberry. – In: Flower morphology and reproductive process of Angiosperms. Moscow, Leningrad. P. 189–204 (In Russ.).
- Su Z., Zhao L., Zhao Y., Li S., Won S., Cai H., Wang L., Li Z., Chen P., Qin Y., Chen X. 2017. The THO complex non-cell-autonomously represses female germline specification through the TAS3-ARF3 module. – Current Biology. 27 (11): 1597–1609.
- Su Z., Wang N., Hou Z., Li B., Li D., Liu Y., Cai H., Qin Y., Chen X. 2020. Regulation of female germline specification via small RNA mobility in *Arabidopsis*. – Plant Cell. 32 (9): 2842–2854.

- Traub M. Sur les Casuarinees et leur place dans le systeme naturel. — *Annales du Jardin Botanique de Buitenzorg*. 10: 145–219.
- Tucker M.R., Koltunow A.M. 2014. Traffic monitors at the cell periphery: the role of cell walls during early female reproductive cell differentiation in plants. — *Current Opinion in Plant Biology*. 17: 137–145.
- Tucker M.R., Okada T., Hu Y., Scholefield A., Taylor J.M., Koltunow A.M. 2012. Somatic small RNA pathways promote the mitotic events of megagametogenesis during female reproductive development in *Arabidopsis*. — *Development*. 139 (8): 1399–1404.
- Tung S.H., Ye X.L., Zee S.Y., Yeung E.C. 2000. The microtubular cytoskeleton during megasporogenesis in the Nun orchid, *Phaius tankervilleae*. — *New Phytologist*. 146 (3): 503–513.
- Twell D. 2011. Male gametogenesis and germline specification in flowering plants. — *Sex. Plant Reprod.* 24 (2): 149–160.
- Van Norman J.M., Breakfield N.W., Benfey P.N. 2011. Intercellular communication during plant development. — *Plant Cell*. 23 (3): 855–864.
- Vasil V. 1959. Morphology and embryology of *Gnetum*. — *Phytomorphology*. 9: 167–215.
- Vinogradova G.Yu. 2009. Poliembrioniya y *Allium ramosum* L. i *A. schoenoprasum* L. (Alliaceae) [Polyembryony in *Allium ramosum* L. and *A. schoenoprasum* L. (Alliaceae)]. — PhD thesis. Saint Petersburg. BIN RAS. 18 p. (In Russ.).
- Vinogradova G.Yu. 2016. Mechanisms of cell differentiation of the female reproductive structures *in vivo* and *in vitro* (for example *Allium* species). — Proceedings of the VII International Scientific and Practical Conference “Biotechnology as an Instrument for Plant Biodiversity Conservation (physiological, biochemical, embryological, genetic and legal aspects)”, devoted to the 30th anniversary of Biotechnology Department in Nikita Botanical Gardens. Simpheropol. 352 p. (In Russ.).
- Vinogradova G.Yu. 2017. Morphogenesis of the female reproductive structures in *Euphorbia* (Euphorbiaceae) species different by the embryo sac development type. — *Bot. Zhurn.* 102 (8): 1060–1093 (In Russ.).
- Vinogradova G.Yu., Zhinkina N.A. 2021. Why does only one embryo sac develop in the *Paeonia* ovule with multiple archesporium? — *Plant Biol.* 23 (2): 267–274.
- Walters J.L. 1962. Megasporogenesis and gametophyte selection in *Paeonia californica*. — *Am. J. Bot.* 49 (7): 787–794.
- Wang N., Huang H.J., Ren S.T., Li J.J., Sun Y., Zhang S.Q. 2012. The rice wall-associated receptor-like kinase gene *OsDEES 1* plays a role in female gametophyte development. — *Plant Physiology*. 160 (2): 696–707.
- Warming E. 1878. De l’ovule. — *Annales des sciences naturelles. Botanique. Series 6*. 5: 177–266.
- Webb M.C., Gunning B.E.S. 1990. Embryo sac development in *Arabidopsis thaliana* — I. Megasporogenesis, including the microtubular cytoskeleton. — *Sex. Plant Reprod.* 3 (4): 244–256.
- Williams C.G. 2009. Conifer reproductive biology. Dordrecht, Heidelberg, London, New York. 174 p.
- Wyrzykowska J., Fleming A. 2003. Cell division pattern influences gene expression in the shoot apical meristem. — *Proceedings of the National Academy of Sciences of the United States of America*. 100 (9): 5561–5566.
- Yakovlev M.S., Yoffe M.D. 1965. Embryology of some representatives of the genus *Paeonia* L. — Flower morphology and reproductive process of Angiosperms. Moscow, Leningrad. P. 140–176.
- Yao X., Yang H., Zhu Y., Xue J., Wang T., Song T., Yang Z., Wang S. 2018. The canonical E2Fs are required for germline development in *Arabidopsis*. — *Frontiers in Plant Science*. 9: 638
- Zhao X., de Palma J., Oane R., Gamuyao R., Luo M., Chaudhury A., Herve P., Xue Q., Bennett J. 2008. OSTDL1A binds to the LRR domain of rice receptor kinase MSP1, and is required to limit sporocyte numbers. — *Plant J.* 54 (3): 375–387.
- Zhao X.A., Bramsiepe J., Van Durme M., Komaki S., Prusicki M.A., Maruyama D., Forner J., Medzihradzsky A., Wijnker E., Harashima H., Lu Y., Schmidt A., Guthörl D., Sahún Logroño R., Guan Y., Pochon G., Grossniklaus U., Laux T., Higashiyama T., Lohmann J.U., Nowack M.K., Schnittger A. 2017. RETINOBLASTOMA RELATED1 mediates germline entry in *Arabidopsis*. — *Science*. 356 (6336): 378–379.
- Zhao L., Liu Y., Dou X., Cai H., Aslam M., Hou Z., Jin X., Li Y., Wang L., Zhao H., Wang X., Sicard A., Qin Y. 2021. Characterization of germline development and identification of genes associated with germline specification in pineapple. — *Horticulture Research*. 8: 239.